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Improvement of model quality by rejection of non-isomorphous frames

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ABSTRACT

Crystallographic data sets consisting of numerous frames usually possess random error, but additionally may display systematic differences arising from radiation damage or the experimental setup. It is crucial to combine only those frames to one complete data set, which are similar and display no systematic differences, which are in fact isomorphous. In this approach a method using $CC_{1/2}$ [1,2] was used to identify non-isomorphous frames from a reference data set [3], reject those and improve the data statistics such as $CC_{1/2}$ and internal correlation of the merged data sets. Moreover, correlation with the previously published model [4] was improved after rejection. CC1/2 therefore correctly predicts non-isomorphism and the agreement of data and model.

167 frames of one out of 5 crystals of the SBDG [3] reference set SNX17 were analysed with XDSGUI (graphical user interface of XDS) (Fig.2) and non-isomorphous frames were identified according to $\Delta CC_{1/2}$ (~>100). Exactly the same tendency can be observed for the correlation of every single frame with the previously published model [4], as non-isomorphous data sets showing negative $\Delta CC_{1/2}$ also show reduced correlation with the model. Rejection of those frames rescaling and merging with 4 other data sets led to improved $CC_{1/2}$, R_{meas} and internal correlation between the merged 5 data sets.

INTRODUCTION

What is non-isomorphism?

A protein crystal is exposed to radiation and the diffraction pattern is used to build the atomic model (Fig.1). During data collection lots of frames from different crystals are measured for reconstruction of the model. Because of radiation damage and different experimental conditions systematic errors, thus non-isomorphism, can occur. Only (similar) isomorphous frames should be merged for calculations.



Why is this so fascinating?



How to find these similar datasets?

METHOD

The $\Delta CC_{1/2}$ method uses $CC_{1/2}$ [1] to identify isomorphous frames. The $CC_{1/2}$ of all frames is calculated which is denoted as **CC**_{1/2_overall}. For every frame $CC_{1/2}$ is again calculated, but one specific frame is excluded during the calculations resulting in $CC_{1/2}$. Finally, the difference is taken:

 $\Delta CC_{1/2} = CC_{1/2 \text{ overall}} - CC_{1/2 \text{ i}}$

Frames with a negative $\Delta CC_{1/2}$ therefore display impairment of the data set and vice versa.

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data 'quality'. Why is this necessary? If data 'quality' is not high enough, the structure can not be solved. Improved 'quality' leads to a solved structure.

Separation of these (somehow) different frames leads to improved

References

separated.

[1] Meyer P. A. et al. (2016). Nat. Commun. 7, 10882. [2] Karplus P. A., Diederichs K. (2012). Science 336, 1030-1.033 [3] Assmann G., Brehm W., Diederichs K. (2016). J.Appl.Cryst. 49, 1021-1028. [4] Stiegler A. L., Zhang R., Liu W., Boggon T. J. (2014). J.Biol.Chem. 289, 25362-25373.